

BOJARSKI HOPPING MODEL OF MULTISTEP NONRADIATIVE ENERGY TRANSPORT. APPLICATION TO SELECTED BIOLOGICAL SYSTEMS

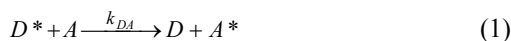
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The applications of Bojarski hopping multistep nonradiative energy transport model to the investigations of energy transfer between biological molecules (for example flavin mononucleotide, FMN) are described. These investigations concern the following problems: (1) the processes of multistep energy migration between biological molecules followed by energy transfer to acceptors, (2) the directional energy transfer in rigid solutions, (3) the manifestations of energy transfer in the system of fluorescent dimers and monomers in rigid matrices containing FMN molecules, (4) the manifestation of reverse energy transport.

INTRODUCTION

Förster resonance energy transfer plays an important role in numerous photochemical processes: photosynthesis, information transfer in DNA, vision process and other photoreception phenomena. During last three decades a lot of papers devoted to resonance energy transfer between biological molecules have been published (Eftink, 1991; Eftink & Ghiron, 1981; Frąckowiak, Cegielski & Abdurakhmanov, 1991 Frąckowiak *et al.*, 1988, 1994; Gobets *et al.*, 2003; Laible *et al.*, 2003; Malicka *et al.*, 2003; Young *et al.*, 2003). Resonance energy transfer has been discovered by Perrin over 80 years ago (Perrin, 1925a; 1925b) and it is still a subject of extensive theoretical and experimental works. In 1949 Förster elaborated a quantum-mechanical theory of single step nonradiative energy transfer from an excited molecule – donor D* to an unexcited acceptor A based on the long range dipole-dipole interaction mechanism (Förster, 1948; 1949). In this case resonance energy transfer can be schematically presented as:



Förster has shown that the probability of energy transfer per time unit can be connected with the transition probability in emission and absorption of interacting molecules. He obtained the expression for the rate constant for nonradiative energy transfer between two transition dipole moments D* and A (Förster, 1948):

$$k^{dd} = \frac{1}{\tau_{0D}} \left(\frac{R_{0DA}}{R} \right)^6 \quad (2)$$

where τ_{0D} is the real averaged lifetime of D* in its excited state when $C_A \rightarrow 0$, R is the distance between D and A, and R_{0DA} is the critical distance for energy transfer (so called Förster radius). R_{0DA} can be determined from the spectroscopic data according to the formula:

$$R_{0DA} = \left(\frac{9 < \kappa^2 > (\ln 10) \eta_{0D} I_{DA}}{128 \pi^5 n^4 N'} \right)^{\frac{1}{6}} \quad (3)$$

where η_{0D} is the donor fluorescence quantum yield for $C_A \rightarrow 0$ is the refractive index of the medium, N' – is the number of molecules in mM, and I_{DA} – the overlap integral:

$$I_{DA} = \int_0^\infty F_D(\bar{\nu}) \varepsilon_A(\bar{\nu}) \bar{\nu}^{-4} d\bar{\nu} \quad (4)$$

where $\varepsilon_A(\bar{\nu})$ is the molar decimal acceptor extinction coefficient, $F_D(\bar{\nu})$ – is the spectral donor fluorescence distribution expressed in the number of quanta and normalized to unity $\int_0^\infty F_D(\bar{\nu}) d\bar{\nu} = 1$, and $\bar{\nu}$ – the wave number (overlap of these spectra is the necessary condition for nonradiative energy transfer). κ^2 in formula (3) is the orientation factor, dependent on mutual orientation of transition dipole moments of interacting D* and A molecules:

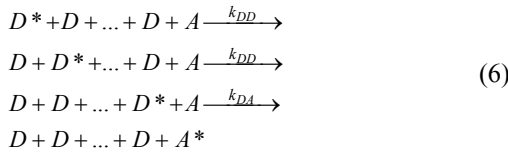
$$\kappa^2 = (\cos \theta_{DA} - 3 \cos \theta_D \cos \theta_A)^2 \quad (5)$$

where θ_{DA} is the angle between two transition moments, θ_D and θ_A are angles between the respective transition moment and the vector connecting D and A molecules.

At the same time other resonance energy transfer theories have also been developed by Dexter, Jabłoński, Galanin, Ore, Knox, Maksimow and Rozman and others. These theories have been limited to the case of low donor concentration relative to the acceptor concentration ($C_D \ll C_A$).

BOJARSKI HOPPING MODEL OF MULTISTEP NONRADIATIVE ENERGY TRANSPORT

In 1971 Bojarski & Domsta (1971) extended the Förster model to arbitrary dye concentration and for the first time took into account the multistep energy migration effect within the donors set on excitation trapping by acceptors. This process can be presented according to the following scheme:



Excitation energy migrates (EM – energy migration) visiting different donor molecules before it is transferred to any acceptor. This is the base for so called hopping forward energy transport model (FET-HM) (Bojarski & Domsta, 1971). This model has then been extended to reverse nonradiative energy transfer (RET-HM) (Bojarski, 1984) from the acceptors set to the donors set. This process takes place if acceptor fluorescence band overlaps partly with the donor absorption band. Then, energy may return to donors according to the scheme:



The nonradiative energy transfer studies require usually the measurements of concentration depolarization and concentration quenching of fluorescence. Bojarski (1984) within the RET-HM obtained the following expression for relative donor quantum yield and emission anisotropy:

$$\frac{\eta_D}{\eta_{0D}} = \frac{1 - f(\gamma)}{1 - \alpha f(\gamma)} \times \frac{1}{1 - B} \quad (8)$$

$$\frac{r_D}{r_{0D}} = (1 - \alpha f(\gamma))(1 - B) \quad (9)$$

$$\text{where } B = \frac{(1 - \alpha)f(\gamma)}{1 - \alpha f(\gamma)} \times \frac{(1 - \alpha')f(\gamma')}{1 - \alpha' f(\gamma')} \quad (10)$$

$$\gamma = \frac{\gamma_{DD}}{\sqrt{2}} + \gamma_{DA} = \frac{\sqrt{\pi}}{2} \left(\frac{C_D}{\sqrt{2}C_{0DD}} + \frac{C_{DA}}{C_{0DA}} \right), \quad (11)$$

$$\alpha = \frac{\gamma_{DD}}{\gamma}$$

$$\gamma' = \frac{\gamma_{AA}}{\sqrt{2}} + \gamma_{DA} = \frac{\sqrt{\pi}}{2} \left(\frac{C_A}{\sqrt{2}C_{0AA}} + \frac{C_D}{C_{0AD}} \right), \quad (12)$$

$$\alpha' = \frac{\gamma_{AA}}{\gamma'}$$

where γ is total reduced concentration in the case of energy migration between donors and nonradiative forward energy transfer from donor to acceptor (FET), γ' – is total reduced concentration for energy migration between acceptors and nonradiative reverse energy transfer from acceptor to donor (RET). Critical concentrations C_{0DD} , C_{0DA} , C_{0AD} and C_{0AA} are given by the formula:

$$C_{0XY} = 4.23 \times 10^{-10} n^2 \left(\kappa^2 > \eta_{0X} I_{XY} \right)^{\frac{1}{2}}, \quad (13)$$

$X, Y \in \{D, A\}$

where I_{XY} is expressed by formula (4). The function $f(\mu)$, where $\mu \in \{\gamma, \gamma'\}$, is connected with the error function $\text{erf}(\mu)$ by:

$$f(\mu) = \sqrt{\pi} \exp(\mu^2) [1 - \text{erf}(\mu)] \quad (14)$$

FET-HM and RET-HM are particularly valuable for studies of energy transfer between biological molecules. In these models energy transfer from donors to acceptors preceded by energy migration between donors can be studied, which takes place in photoreception systems or photosynthesis phenomena. Fluorescence concentration quenching (FCQ) model elaborated by Bojarski can also be used to study energy transport if the role of acceptors is played by dimers (Bojarski & Kowski, 1986; Bojarski & Sienicki, 1990).

APPLICATION OF BOJARSKI MODEL TO BIOLOGICAL MOLECULES

In rigid systems (PVA films or biological membranes) the directional energy transfer can take place (Bojarski & Sienicki, 1990; Grajek, 2007; Grajek *et al.*, 1998; Żurkowska, Grajek & Bojarski, 1996) due to inhomogeneous orientation broadening of energy levels (Rubinov, Tomin & Bushuk, 1982; Yuzhakov, Grachev & Ponomarev, 1990). This effect leads to the formation of energetic continuous distribution of fluorescent centers with different energies of 0–0 transitions. Additionally, if dimers appear in the system, then the energy from excited monomer centers flows to energetically lower

dimeric centers (Grajek, 2007). In photosynthetic systems such a directional energy transfer is important, since in reaction centers, in which the photochemical reaction takes place, dimers appear and monomers being the antennas transfer the absorbed energy to such centers. Moreover, in rigid systems dimers exhibit weak fluorescence and are able to return energy to monomers (Bojarski *et al.*, 2003; Grajek, 2007).

In this work based on Bojarski model all processes of energy transport between fluorescent flavin mononucleotide (FMN) monomers (donors) and dimers (acceptors) in PVA film can be analyzed.

FMN is a biologically important molecule acting as coenzyme and photoreceptor in living organisms. FMN acts as a photoreceptor in many plants (Kottke *et al.*, 2003; Okajima *et al.*, 2006). Recent studies have shown the existence of flavins dimers in biological systems (Bieger, Essen & Oesterheld, 2003; Muralidhara & Wittung-Stafshede, 2003) as well as in photoreception systems (Nakasako *et al.*, 2005; Salomon, Lempert & Rüdiger, 2004). Salomon, Lempert & Rüdiger (2004) have shown that the fragments of phototropin – photoreceptor containing LOV domains with FMN act as dimers. Detailed knowledge of the system studied is important for understanding the processes occurring in biological systems containing FMN.

The dimerization of FMN in water (Grajek *et al.*, 1984, 2001), glycerol-water solutions (Grajek *et al.*, 1984) and solid PVA matrices (Grajek, 2003; 2007; Grajek *et al.*, 1998) has been studied in detail. Investigations performed in (Grajek, 2007; Grajek *et al.*, 1986) have shown the existence of dimer electronic level splitting to H and J levels (Kasha, Rawls & El-Bayoumi, 1965). It has been shown that both levels can be active in nonradiative energy transfer in rigid systems (Grajek, 2007).

FMN is a very interesting and exceptional model system for study of energy transfer because it practically does not form aggregates of higher order than dimers (Grajek, 2003; 2007; Grajek *et al.*, 1986; 2001, 2007) and FMN dimers in rigid solution have also a unique feature – they are fluorescent (Grajek, 2007; Grajek *et al.*, 1998, 2007) and form imperfect traps for excitation energy (Bojarski *et al.*, 2003; Grajek, 2007).

Fluorescence (Grajek, 2007; Grajek *et al.*, 1998; Żurkowska, Grajek & Bojarski, 1996) and fluorescence anisotropy (Grajek, 2007) measurements of FMN in PVA versus concentration at different excitation and observation wavelengths allowed for directional energy migration from “blue” to “red” centers to be observed. These observations were possible both for monomers and dimers (Grajek, 2007). Also directional energy transfer from monomers to dimers, reverse energy transfer from dimers to monomers and remigration to primarily excited monomers could be analyzed (Grajek, 2007). The results obtained (Grajek, 2007) have shown that the

excitation wavelength is the most important factor determining the nonradiative energy transport processes and allowing for their control.

Measurements of fluorescence or fluorescence anisotropy spectra upon different excitation wavelength enable to observe energy migration tracking between donors (monomers) which took place more or less effectively depending on the excitation wavelength. Excitation wavelength controlled the number of excitation transfer acts and the whole process of multistep energy transfer within the set of FMN monomers and dimers (Grajek, 2007), and allowed for the observation of energy transfer with the contribution of dimeric H and J levels (Grajek, 2007). For example, at $\lambda_{\text{ex}} = 445$ nm (maximum of monomer emission and maximum of H dimer absorption) numerous processes of possible energy transport acts are possible in the set of FMN monomers and dimers in PVA ($M^* \rightarrow M$, $M^* \rightarrow D$, $D^* \rightarrow M$, $D^* \rightarrow D$) in which H and J dimer levels can participate. Hence, low emission anisotropy values were obtained for such conditions (Jabłoński, 1970; 1972). However, upon the excitation $\lambda_{\text{ex}} = 510$ nm (dimers are excited to the J level) only energy migration was observed between dimers.

It should be mentioned that depending on the selected observation wavelength more or less depolarized FMN fluorescence was also observed due to stronger or weaker sink of multistep energy migration. Investigations at different observation wavelengths enable tracking of particular stages of multistep nonradiative energy transport by observation of light originating from given centers starting from the “blue” ones and ending at the “red” ones (Grajek, 2007). For example, emission anisotropy measurements carried out for extremely concentrated samples, $C = 0.684$ M and $C = 1$ M, at $\lambda_{\text{obs}} = 510$ nm allowed to observe fluorescence originating from short wavelength dimeric centers and primarily excited molecules which resulted in high emission anisotropy values (Grajek, 2007), whereas at $\lambda_{\text{obs}} = 600$ nm depolarized fluorescence was observed due to energy transfer to long wavelength centers.

Also the quantitative studies of resonance energy transfer with the use of FET-HM and RET-HM (Eqs. 8 and 9) as well as Monte-Carlo simulations were performed for the concentration quenching and depolarization of FMN fluorescence (Grajek, 2007). These studies were carried out using different models too (Bojarski *et al.*, 2003). The obtained agreement between the theoretical and experimental data confirmed the correctness of the analysis of energy transport processes in the system of interest. Theoretical data (Grajek, 2007) confirmed: (1) the existence of fluorescent FMN dimers and their contribution to the resonance energy transfer (2) the appearance of multistep nonradiative energy transport, (3) existence of reverse energy transfer from D^* to M, (4) high role of energy migration in the monomer and

dimer sets. The calculated values of overlap integrals for nonradiative energy transport I_{XY} from formula (4), where $X, Y \in \{D, A\}$, have shown that the probability of energy migration between dimers is higher than that from dimers to monomers of FMN in PVA. In rigid systems, where fluorescent dimers are formed the return of the excitation energy from D^* to M is possible and the energy transport is well described by the RET-HM. In the case of high temperatures where the FMN dimers in PVA become perfect traps the energy transport is well described by FET-HM. FET-HM has been successfully applied to describe multistep nonradiative energy transport from excited monomers M^* to dimers – energy traps in the case of glycerol – aqueous FMN solutions (Grajek *et al.*, 1992). However, in the case of aqueous solutions the FET-HM takes into account the effect of diffusion on resonance energy transfer (Grajek, Żurkowska & Kuśba, 2005).

The models mentioned are widely used for studies of different donor-acceptor systems, both fluid and viscous or rigid (Bojarski, 1997a; Bojarski & Kułak, 1997; 1998a) or in the case of rhodamine dimers (Bojarski, 1997b; Bojarski & Kułak, 1998b; Bojarski *et al.*, 1996) as well as in partly ordered systems (Bojarski *et al.*, 2003, 2004; Bojarski, Kułak & Kamińska, 2002; Sadownik *et al.*, 2007).

Taking account of the diversity of energy transport processes (Grajek, 2007) the question arises: Do living organisms select certain energy quanta out of the solar spectrum to excite photoreceptors which in turn activate only specific processes of energy transfer?

Abbreviations: EM – energy migration, FET-HM – forward energy transfer hopping model, RET-HM – reverse energy transfer hopping model, FET – forward energy transfer, RET – reverse energy transfer, FCQ – fluorescence concentration quenching

REFERENCES

- Bieger B., Essen L.O. & Oesterhelt D. (2003). Crystal structure of halophilic dodecin: a novel, dodecameric flavin binding protein from *Halobacterium salinarum*. *Structure*, **11**, 375–385.
- Bojarski C. (1984). Influence of the reversible energy transfer on the donor fluorescence quantum yield in donor-acceptor systems. *Z. Naturforsch. Teil A*, **39**, 948–951.
- Bojarski C. & Domsta J. (1971). Theory of the influence of concentration on the luminescence in solid solutions. *Acta Phys. Acad. Sci. Hung.*, **30**, 145–166.
- Bojarski C. & Kowski A. (1986). *Międzymolekularne oddziaływania w stanach wzbudzonych*. Zakład Narodowy Imienia Ossolińskich, PAN, Wrocław, s. 41–109.
- Bojarski C. & Sienicki K. (1990). Energy transfer and Migration in Fluorescent Solutions. [In:] *Photochemistry and Photophysics*, Vol. 1. ed. J.F. Rabek. CRC Press, Inc. Boca Raton, Florida.
- Bojarski P. (1997a). Temperature effect on the fluorescent spectra of two-component viscous solutions in the presence of forward and reverse nonradiative excitation energy transport. *J. Luminescence*, **71**, 47–56.
- Bojarski P. (1997b). Concentration quenching and depolarization of rhodamine 6G in the presence of fluorescent dimers in polyvinyl alcohol films. *Chem. Phys. Letters*, **278**, 225–232.
- Bojarski P., Kułak L., Grajek H., Żurkowska G., Kamińska A., Kukliński B. & Bojarski C. (2003). Excitation energy transport and trapping in concentrated solid solutions of flavomononucleotide. *Biophys. Biochim. Acta*, **1619**, 201–208.
- Bojarski P. & Kułak L. (1997). Forward and reverse nonradiative excitation energy transport in concentrated two-component systems. *Chem. Phys.*, **220**, 323–336.
- Bojarski P. & Kułak L. (1998a). Excitation energy transport between the ionic forms of rhodamine B in viscous solutions. *Acta Phys. Polon.*, **94**(5–6), 725–734.
- Bojarski P. & Kułak L. (1998b). Nonradiative excitation energy transport between monomers and fluorescent dimers of rhodamine 6G in polyvinyl alcohol films. *Asian J. Spectroscopy*, **2**, 91–102.
- Bojarski P., Gryczyński I., Kułak L., Synak A. & Barnett A. (2006). Excitation energy migration between elongated fluorophores in uniaxially oriented polyvinyl alcohol films. *Chemical Physics Letters*, **431**, 94–99.
- Bojarski P., Kamińska A., Kułak L. & Sadownik M. (2003). Excitation energy migration in uniaxially oriented polymer films. *Chem. Phys. Lett.*, **375**, 547–552.
- Bojarski P., Kułak L. & Kamińska A. (2002). Nonradiative excitation energy transport and its analysis in concentrated systems. *Am. J. Spectroscopy*, **5**, 145–163.
- Bojarski P., Mateczuk A., Bojarski C., Kowski A., Kukliński B., Żurkowska G. & Diehl H. (1996). Fluorescent dimers of rhodamine 6G in concentrated ethylene glycol solution. *Chem. Phys.*, **210**, 485–499.
- Eftink M.R. (1991). Fluorescence quenching reactions. Probing biological macromolecular structures. [In:] *Biophysical and biochemical aspects of fluorescence spectroscopy*. ed. T. G. Dwey, Plenum Press, New York, 1–41.
- Eftink M.R. & Ghiron C.A. (1981). Fluorescence quenching studies with proteins. *Anal. Biochem.*, **114**, 199–227.
- Förster T.H. (1948). Zwischenmolekulare Energiewanderung und Fluoreszenz. *Ann. Phys. (Leipzig)*, **2**, 55.
- Förster T.H. (1949). Experimentelle und theoretische Untersuchung des zwischenmolekularen Übergangs von Elektronenanregungsenergie. *Z. Naturforsch. Teil A*, **4**, 321–327.
- Frąckowiak D., Cegielski R. & Abdurakhmanov I.A. (1991). Excitation energy transfer from carotenoids to bacteriochlorophyll in the bacterium *Chromatium minutissimum*. *Photosynthetica*, **25**, 621–629.
- Frąckowiak D., Romanowski M., Hotchandani S., Leblanc L. & Leblanc R.M. (1988). A photoelectrochemical cell with oriented pigment-polymer complexes. *Bioelectrochem. Bioenerg.*, **19**, 371–387.
- Frąckowiak D., Zelnet B., Malak H., Cegielski R., Planner A., Goc J. & Niedbalska N. (1994). Path of deactivation of excitation of *Chlorophyll a* in various model systems. *Proc. SPIE*, **2137**, 643–660.
- Gobets B., Stokkum I.H.M., Mourik F., Dekker J.P. & Grondelle R. (2003). Excitation wavelength dependence of

- the fluorescence kinetics in photosystem I particles from *Synechocystis* PCC 6803 and *Synechococcus elongatus*. *Biophys. J.*, **85**, 3883–3898.
- Grajek H. (2003). The effect of temperature on FMN absorption spectra in rigid poly(vinyl alcohol) matrices. *Biochim. Biophys. Acta*, **1620**, 133–138.
- Grajek H. (2007). *Procesy bezpromienistego przekazywania elektronowej energii wzbudzenia między monomerami i fluoryzującymi dimerami mononukleotydu flavinowego – FMN w układach sztywnych*. Rozprawy i monografie (Dissertations and monographs). Wydawnictwo Uniwersytetu Warmińskiego-Mazurskiego, pp. 3–158.
- Grajek H., Drabent R., Żurkowska G. & Bojarski C. (1984). Absorption of the flavin dimers. *Biochim. Biophys. Acta*, **801**, 456–460.
- Grajek H., Gryczyński I., Bojarski P., Gryczyński Z., Bharill S. & Kuślak L. (2007a). Flavinmononucleotide fluorescence intensity decay in concentrated aqueous solutions. *Chem. Phys. Lett.*, **439**, 151–156.
- Grajek H., Liwo A., Wiczak W. & Żurkowska G. (2007b). Resolution of the excitation-emission spectra of FMN in rigid poly(vinyl alcohol) matrices. *J. Photochem. Photobiol B*, **86**, 193–198.
- Grajek H., Żurkowska G. & Kuśba J. (2005). Influence of diffusion on nonradiative energy transfer between FMN molecules in aqueous solutions. *Biochim. Biophys. Acta*, **80(2)**, 145–155.
- Grajek H., Bojarski C., Żurkowska G. & Drabent R. (1992). The influence of fluorescence concentration quenching on the emission anisotropy of flavins in glycerine-water solutions. *Photochem. Photobiol.*, **55**, 381–387.
- Grajek H., Żurkowska G., Bojarski P., Kukliński B., Smyk B., Drabent R. & Bojarski C. (1998). Spectroscopic manifestations of flavomononucleotide dimers in polyvinyl alcohol films. *Biochim. Biophys. Acta*, **1384**, 253–267.
- Grajek H., Żurkowska G., Drabent R. & Bojarski C. (1986). The structure of the flavomononucleotide dimer. *Biochim. Biophys. Acta*, **881**, 241–247.
- Grajek H., Żurkowska G., Drabent R. & Bojarski C. (2001). The effect of temperature and concentration on the aggregation of flavomononucleotide in aqueous solutions. *Asian J. Spectroscopy*, **2**, 49–55.
- Jabłoński A. (1970). Anisotropy of fluorescence of molecules excited by excitation transfer. *Acta Phys. Polon.*, **A38**, 453–458.
- Jabłoński A. (1972). Depolarization of fluorescence of isotropic solutions produced by excitation transfer between luminescent molecules. *Acta Phys. Polon.*, **A41**, 85–90.
- Kasha M., Rawls H.R. & El-Bayoumi M.A. (1965). The excitation model in molecular Spectroscopy. *Pure Appl. Chem.*, **11**, 371–392.
- Kottke T., Heberle J., Hehn D., Dick B. & Hegemann P. (2003). Phot-LOV1: Photocycle of a blue-light receptor domain from the green *Alga Chlamydomonas reinhardtii*. *Biophys. J.*, **84**, 1192–1201.
- Laible P.D., Morris Z.S., Thurnauer M.C., Schiffer M. & Hanson D.K. (2003). Inter- and intraspecific variation in excited-state triplet energy transfer rates in reaction centers of photosynthetic bacteria. *Photochem. Photobiol.*, **78(2)**, 114–123.
- Malicka J., Gryczyński I., Fang J., Kuśba J. & Lakowicz J.R. (2003). Increased resonance energy transfer between fluorophores bound to DNA in proximity to metallic silver particles. *Anal. Biochem.*, **315(2)**, 160–169.
- Muralidhara B.K. & Wittung-Stafshede P. (2003). Can cofactor-binding sites in proteins be flexible? *Desulfovibrio desulfuricans*. flavodoxin binds FMN dimer. *Biochemistry*, **42**, 13074–13080.
- Nakasako M., Matsuoka D., Zikihara K. & Tokutomi S. (2005). Quaternary structure of LOV-domain containing polypeptide of *Arabidopsis* FKF1 protein. *FEBS Lett.*, **579(5)**, 1067–1071.
- Okajima K., Fukushima Y., Suzuki H., Kita A., Ochiai Y., Katayama M., Shibata Y., Miki K., Noguchi T., Itoh S. & Ikeuchi M. (2006). Fate determination of the flavin photoreceptors in the cyanobacterial blue receptor TePixD (TH0078). *J. Mol. Biol.*, **363**, 10–18.
- Perrin J. (1925a). *2^{me} Conseil de Chim.* Solvay. Gauthier-Villars, Paris, 332.
- Perrin J. (1925b). Sur le mouvement brownien de rotation. *C.R. hebdom.*, *Seances Acad. Sci.*, **181**, 514.
- Rubinow A.N., Tomin V.I. & Bushuk B.A. (1982). Kinetic spectroscopy of orientational states of solvated dye molecules in polar solutions. *J. Lumin.*, **26**, 377–391.
- Sadownik M. & Bojarski P. (2004). The effect of intermolecular donor – acceptor energy transfer on emission anisotropy in uniaxially oriented polymer films. *Chem. Phys. Letters*, **396**, 293–297.
- Sadownik M., Bojarski P., Kwiec P. & Rangelowa S. (2007). Energy transfer between unlike fluorophores in uniaxially oriented polymer films monitored by time – resolved and steady – state emission anisotropy. *Optical Materials*, (after proof).
- Salomon M., Lempert U. & Rüdiger W. (2004). Dimerization of the plant photoreceptor phototropin is probably mediated by the LOV1 domain. *FEBS Lett.*, **572(1–3)**, 8–10.
- Young M., Damjanovic A., Vaswani H.M., & Fleming G.R. (2003). Energy transfer in Photosystem I of cyanobacteria *Synechococcus elongatus*: model study structure-based semi-empirical hamiltonian and experimental spectral density. *Biophys. J.*, **85**, 140–158.
- Yuzhakov V.I., Grachev A.V. & Ponomarev A.N. (1990). Spectroscopic manifestation of association of rhodamine dye molecules in polymer matrices. *Opt. Spektrosk.*, **68**, 39–62.
- Żurkowska G., Grajek H. & Bojarski C. (1996). Effect of concentration and excitation wavelength on fluorescent spectra of flavomononucleotide in polyvinyl alcohol films. *Current Topics in Biophys.*, **20(2)**, 134–138.